

REVIEW

Open Access



Unraveling neuronal and metabolic alterations in neurofibromatosis type 1

Valentina Botero^{1,6,7} and Seth M. Tomchik^{1,2,3,4,5,6*} 

Abstract

Neurofibromatosis type 1 (OMIM 162200) affects ~1 in 3,000 individuals worldwide and is one of the most common monogenetic neurogenetic disorders that impacts brain function. The disorder affects various organ systems, including the central nervous system, resulting in a spectrum of clinical manifestations. Significant progress has been made in understanding the disorder's pathophysiology, yet gaps persist in understanding how the complex signaling and systemic interactions affect the disorder. Two features of the disorder are alterations in neuronal function and metabolism, and emerging evidence suggests a potential relationship between them. This review summarizes neurofibromatosis type 1 features and recent research findings on disease mechanisms, with an emphasis on neuronal and metabolic features.

Keywords Neurofibromatosis type 1, Neurofibromin, NF1, Metabolism

Introduction

Neurofibromatosis type 1 (NF1) is a multisystemic autosomal-dominant condition affecting ~1 in 3,000 live births worldwide [1–4]. Historical recognition of the disorder dates to ancient Egypt [5, 6], with Friedrich Daniel von Recklinghausen providing the first comprehensive clinical description in 1882. Initially termed “von Recklinghausen disease,” it is now known as neurofibromatosis type 1 [7, 8]. Significant progress has been made in understanding NF1's genetic underpinnings. The

disorder is caused by mutations in a single gene, neurofibromin 1 (*NF1*) [9], which encodes the protein neurofibromin (Nf1). Several pivotal discoveries made in the late 1990s include the establishment of diagnostic criteria for accurate NF1 diagnosis [10, 11], mapping of the *NF1* gene to chromosome 17q11.2 [12–14], and the identification of its protein product [12, 15–19]. Despite these advances (and more since then), gaps persist in our understanding of the disorder and its underlying genetic, cellular, and systemic mechanisms. Features of the disorder include alterations in neuronal and brain function as well as metabolic alterations [20–24]. Some of the features of the disorder, including the brain/cognitive symptoms, could be influenced by the metabolic alterations. Here we review the mechanisms of NF1 pathophysiology, with a focus on the emerging understanding of neuronal and metabolic alterations.

Diagnostic criteria and clinical features

NF1 is characterized by a broad spectrum of clinical manifestations that begin in infancy and progressively worsen (Fig. 1) [25, 26]. Diagnostic criteria for NF1, first established in 1987 [10] and updated in 2021 [11] (Table 1), rely on a physical examination and

*Correspondence:

Seth M. Tomchik
seth-tomchik@uiowa.edu

¹ Department of Neuroscience and Pharmacology, University of Iowa, Iowa City, IA, USA

² Stead Family Department of Pediatrics, University of Iowa, Iowa City, IA 52242, USA

³ Iowa Neuroscience Institute, University of Iowa, Iowa City, IA 52242, USA

⁴ Fraternal Order of Eagles Diabetes Research Center, University of Iowa, Iowa City, IA 52242, USA

⁵ Hawk-IDDRC, University of Iowa, Iowa City, IA 52242, USA

⁶ Department of Neuroscience, Scripps Research, Scripps Florida, Jupiter, FL, USA

⁷ Skaggs School of Chemical and Biological Sciences, Scripps Research, La Jolla, CA, USA



© The Author(s) 2024. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

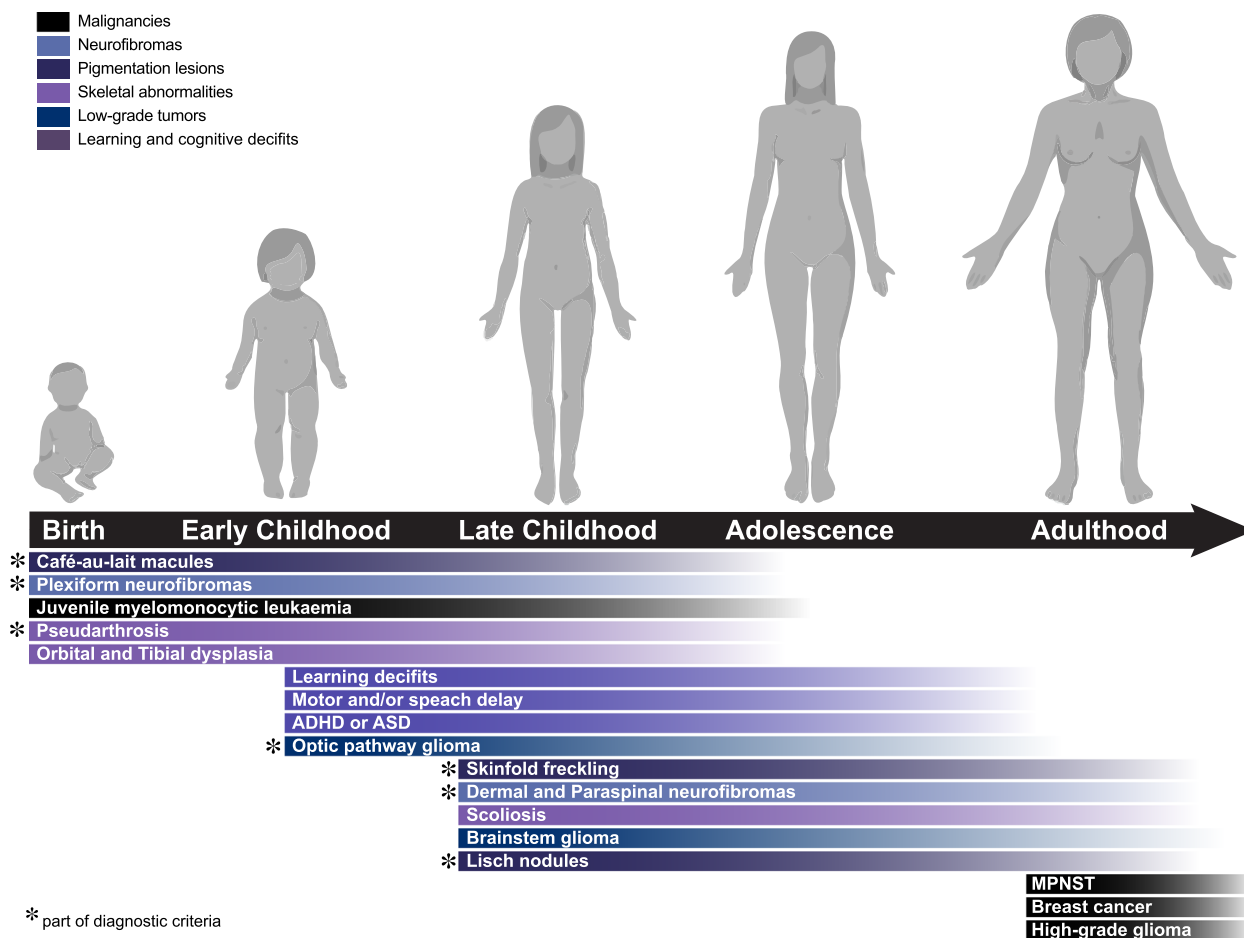


Fig. 1 Disease progression and clinical features of NF1. The onset and severity of NF1 clinical features vary between individuals. In children, the most common clinical physical manifestations are skeletal abnormalities such as scoliosis, tibial dysplasia, and café-au-lait spots. Young children are at risk of developing juvenile myelomonocytic leukemia, optic gliomas, and behavioral and cognitive deficits, with attention-deficit/hyperactivity disorder (ADHD) and autism spectrum disorder (ASD) being the most common. The risk of developing plexiform neurofibromas (pNF) is high during the early stages of life, but other malignancies such as malignant peripheral nerve sheath tumors (MPNST) and breast cancer occur more often after the third decade of life [25, 26]. Created with BioRender.com

Table 1 Diagnostic criteria for NF1

Diagnostic Criteria

Individual presents with two^a or more of the following:

1. Six or more café-au-lait macules of ≥ 5 mm in diameter before puberty or ≥ 15 mm in diameter after puberty
2. Axillary or inguinal freckling
3. Two or more dermal neurofibromas or one plexiform neurofibroma
4. An optic pathway glioma
5. Two or more iris Lisch nodules or choroidal abnormalities
6. A distinctive osseous lesion such as a sphenoid dysplasia, anterolateral bowing of the tibia, or pseudoarthritis of a long bone
7. A heterozygous NF1 variant fraction of 50% in apparently normal tissue such white blood cells

^a A child of a parent with NF1 merits diagnosis if one or more of the features are present

family history review. Some symptoms emerge in an age-dependent manner, making proper diagnosis during early childhood challenging, particularly for those lacking a

family history of the disease (half of NF1 cases stem from de novo mutations) [3, 11, 20]. These challenges led to revisions of the diagnostic criteria, incorporating mosaic

neurofibromatosis and genetic testing [11]. Symptoms of NF1 include increased susceptibility to various tumors, including peripheral nerve tumors like neurofibromas, plexiform neurofibromas, and malignant peripheral nerve sheath tumors, as well as brain tumors such as optic pathway gliomas and brainstem gliomas [1, 25, 27, 28]. Although tumors are a primary clinical characteristic of NF1, it also produces non-tumor symptoms including pigmentation defects, skeletal abnormalities, stunted growth, cognitive impairments, and behavioral alterations [27, 29]. NF1 reduces life expectancy by 8–15 years [30–32] and significantly impacts quality of life, with up to 80% of children experiencing moderate to severe cognitive impairments [22, 29].

The NF1 gene and neurofibromin protein

NF1 results from mutations in the *NF1* gene, which encodes a large 2,818 amino acid protein called neurofibromin (Nf1) [17, 33]. The Nf1 protein contains a central Ras-GTPase activating protein (GAP)-related domain (GRD) [16, 34]. It primarily localizes to the cytoplasm, interacting with Ras at the plasma membrane, and is also found in the nucleus, endoplasmic reticulum, and mitochondria [35–38]. Nf1 is ubiquitously expressed throughout development, with the highest levels in nervous system cells, including Schwann cells, neurons, astrocytes, and oligodendrocytes [17, 19, 39]. Clinical

manifestations of NF1 are variable, even among identical germline *NF1* mutations, and some exhibit segmental or mosaic NF1.

Role of the NF1 gene and neurofibromin GAP-related domain

Over 2,600 unique mutations within the *NF1* gene have been identified [40] (Fig. 2). Clinical heterogeneity of NF1 can be attributed to multiple factors, including allelic variation, second-hit mutations, epigenetic changes, differences across the *NF1* mutations, and tissue-specific Nf1 functions [41]. Nf1 protein expression across different mutations (i.e., Nf1 heterozygosity can result in ~12–89% of normal protein expression level) [42]. Neurofibromas result from loss of *NF1* heterozygosity following second-hit mutations [43]. Other symptoms of the disorder emanate from haploinsufficiency due to the heterozygous mutation itself. Heterozygous germline mutation in *NF1* is associated with notable impacts on cognitive functions, affecting attention and learning and increasing the prevalence of autism spectrum disorder (ASD) [44, 45]. Among the *NF1* isoforms, certain variants have a tissue-specific role. The alternatively-spliced exon 11alt12 (formerly known exon 9a) is predominantly expressed in the central nervous system (CNS), particularly within forebrain neurons [46]. In contrast, the alternatively spliced exon 30alt31 (formerly known as exon

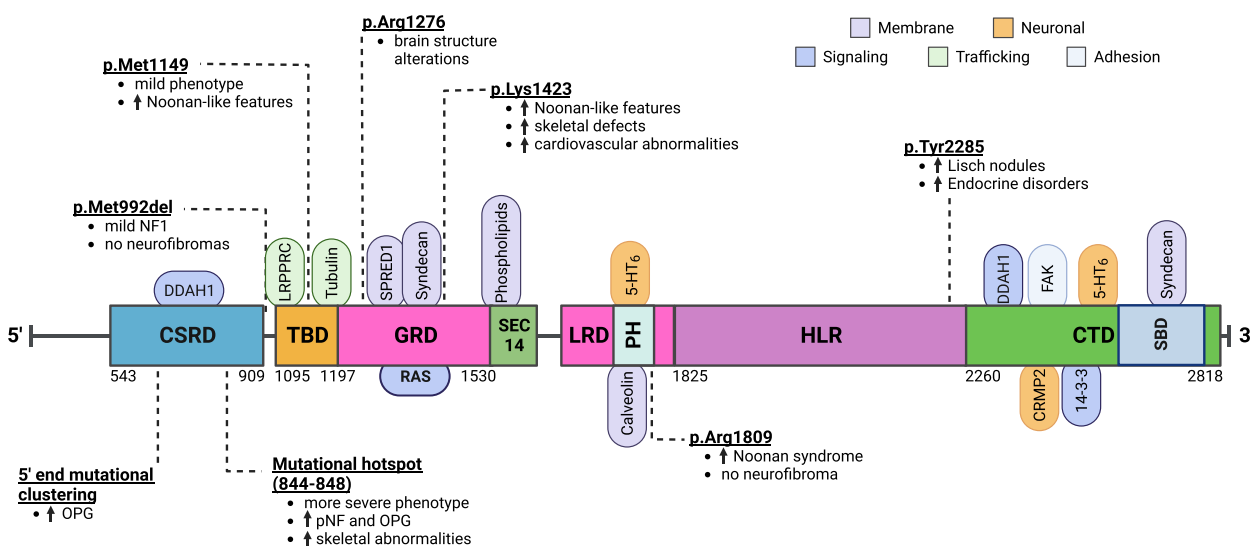


Fig. 2 Nf1 protein structure, interacting domains, and genotype–phenotype correlations. Nf1 protein contains several domains (squares) and interacting proteins (ovals). Nf1 protein domains include the following: cysteine-serine-rich domain (CSRD), tubulin binding domain (TBD), central GTPase-activating-protein-related domain (GRD), SEC14 domain, leucine-rich domain (LRD), pleckstrin homolog (PH), HEAT-like repeats (HLR), C-terminal domain (CTD), syndecan-binding domain (SBD). Phospholipids and proteins identified as Nf1-interacting proteins are shown in association with their described function, such as: trafficking (green), neuronal (yellow), membrane localization (purple), cell adhesion (gray), and cell signaling (blue). Nf1 mutations reported to correlate with certain phenotypes are shown above/below the protein and associated phenotypes. Numbers along protein indicate amino acid residues. Figure adapted and modified from Ratner and Miller (2015) [26], Mo et al. (2022) [59], and Anastasaki et al. (2022) [60]. Created with BioRender.com

23a) contains an alternative exon that lies within the GRD and diminishes Ras GAP activity [47]; mice lacking this exon have learning and communication impairments but are not susceptible to tumor formation [48–50]. Alternative splicing at the 3' end of *NF1* produces the alternative exon 56alt57 (formerly 48a) which is expressed highly in fetal and adult cardiac and skeletal tissue and may contribute to reduced muscle strength and muscle weight [47, 51–54]. Exon 12alt13 (formerly known as 10a-2) is a low-level ubiquitous isoform concentrated in perinuclear granular structures [55]. Moreover, the *NF1*- Δ E43 isoform shows elevated expression in the liver, kidneys, lungs, placenta, and skeletal muscle relative to the general expression of *NF1* [56, 57]. Dimerization of the Nf1 protein and the varied impacts of different mutations on protein stability further complicate the disease's pathophysiology [58].

Neurofibromin's impact on cellular processes via Ras and cAMP signaling

The Nf1 protein features multiple structural domains, with the GRD being the most extensively studied [16, 59–62]. The GRD plays a pivotal role in the regulation of Ras signaling, catalyzing hydrolysis of Ras-bound GTP into GDP and thereby attenuating Ras signaling (Fig. 3). Consequently, *NF1* loss-of-function mutations lead to the accumulation of active Ras-GTP and aberrant activation of downstream pathways, including Raf/MEK/ERK and PI3K/AKT/mTOR [45, 61, 63, 64]. Numerous mutations that compromise the function of the GRD have been identified in patients (Fig. 3) [61]. Analysis of the crystallographic structure of the Nf1 GRD revealed a critical arginine finger residue (R1276) that stabilizes and positions Ras association with the catalytic domain. Notably, a patient mutation (R1276P), which substitutes arginine with proline, results in a >1000-fold reduction in Ras-GAP activity [61, 62, 65].

Nf1 regulates multiple cellular processes, including metabolism, cell proliferation, differentiation, and survival via its regulatory effects on Ras signaling. Nf1/Ras activity is regulated by upstream signal transduction pathways involving receptor tyrosine kinases (RTK). One such RTK is the Anaplastic Lymphoma Kinase (ALK), which interacts with Nf1 and functions as an upstream activator of Nf1-regulated Ras signaling pathway [67–70]. In addition to RTKs, other upstream regulators of Nf1/Ras include G protein-coupled receptors (GPCRs), specifically the G $\beta\gamma$ subunits that bind to Nf1 in striatal neurons and inhibits its capacity to suppress Ras/AKT/mTOR signaling [71].

Loss-of-function mutations in Nf1 dysregulate multiple signaling pathways downstream of Ras, including the canonical mitogen-activated protein kinase (MAPK)

signaling pathway (Raf/MEK/ERK), PI3K/AKT/mTOR, and others (Fig. 3) [18, 63, 64]. These pathways in turn regulate multiple cellular and metabolic processes, including cell growth, survival, nutrient uptake, proliferation, and the modulation of neuronal metabolism in response to growth factors, nutrients, and changes in the cellular energy state [72]. Hyperactivation of Raf/MEK/ERK due to loss of Nf1 is one of the major mechanisms implicated in NF1 phenotypes and is a current therapeutic target [73–75]. Conditional Nf1 knockout in various brain cells (astrocytes, pyramidal cells, GABAergic neurons, and inhibitory/excitatory neurons) increases ERK signaling [37]. In addition, hyperactivation of the PI3K/AKT/mTOR pathway contributes to the development of some NF1-associated phenotypes. The convergence of these two Ras effector pathways, each of which will be discussed in greater detail below, underscores the complexity of cellular signaling alterations in NF1. Overall, the NF1 gene and the Nf1 GRD function as a central regulator of Ras signaling, modulating downstream signaling targets and affecting diverse cellular functions.

In addition to its role in regulating Ras signaling, Nf1 is a positive regulator of cyclic adenosine monophosphate (cAMP) and downstream protein kinase A (PKA) activity (Fig. 3). Nf1 is required for normal cAMP generation in neurons and astrocytes, as observed in *Drosophila* and rodent models of NF1 [76, 77]. In turn, alterations in cAMP/PKA signaling are implicated in many of the NF1 phenotypes, including cell differentiation/growth and learning [68, 78–81]. Furthermore, NF1-related changes in cAMP/PKA levels are altered via a non-canonical mechanism involving Ras-dependent phosphorylation of protein kinase C zeta (PKC ζ) [82, 83]. This pathway regulates neuronal cAMP homeostasis in both human-induced pluripotent stem cell (hiPSC)-derived neurons and primary mouse neuron cultures [82].

Mechanisms of pathophysiology in neurofibromatosis type 1

Tumors

Tumor formation is a primary concern in NF1, with cutaneous neurofibromas (CN) and plexiform neurofibromas (pNF) pervasive among patients. CNs, affecting over 99% of NF1-afflicted adults, are benign but prolific tumors that emerge during late childhood and experience rapid growth during puberty and pregnancy [4]. The number of CNs in adults can reach into the thousands, leading to significant disfigurement and considerable physical and psychological distress [84, 85]. Conversely, pNFs affect about 50% of patients, proliferating rapidly during childhood and adolescence [86]. These tumors are located in peripheral nerve sheaths, with Schwann cells representing the major neoplastic cell type [87]. Neurofibromas

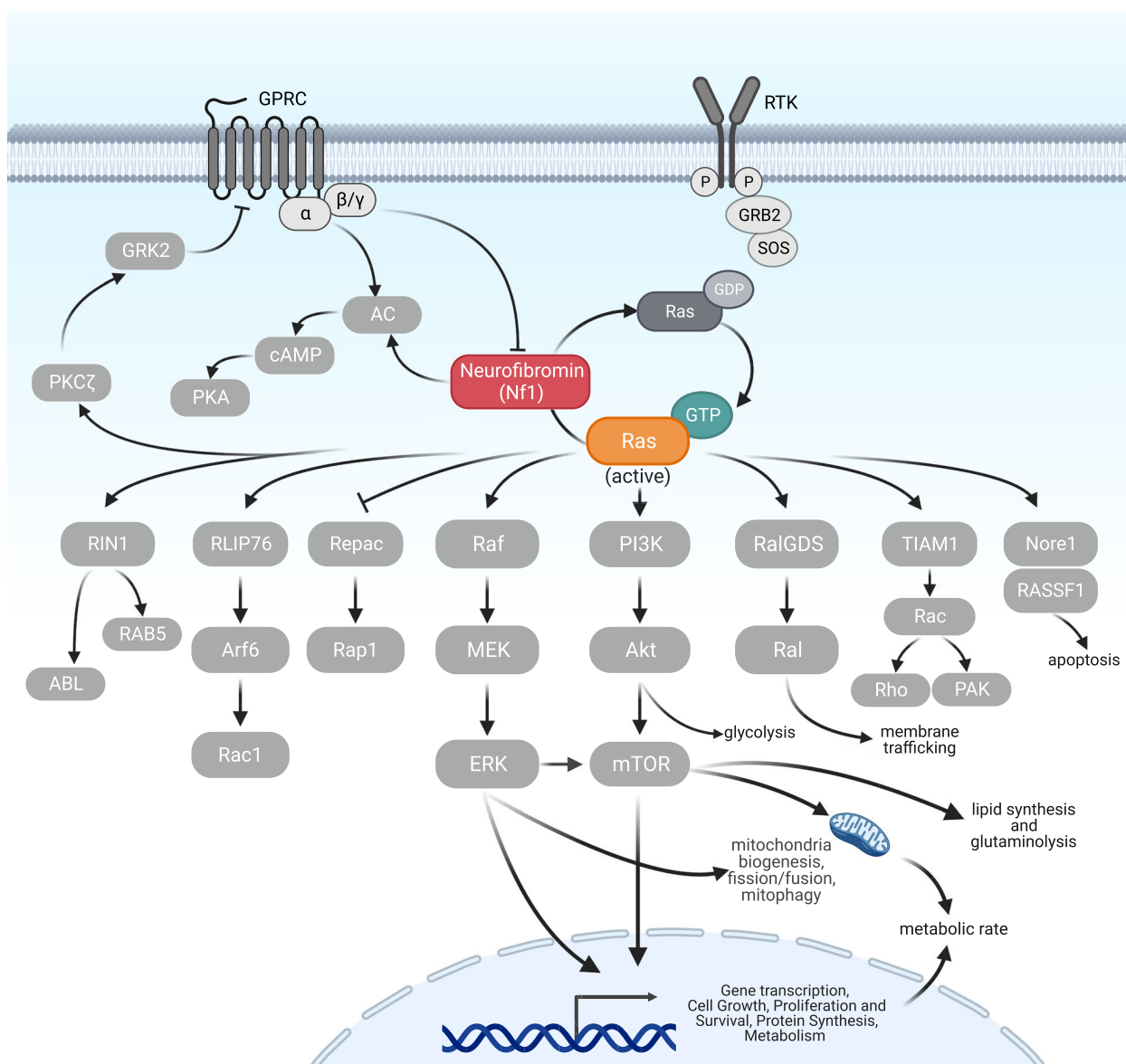


Fig. 3 Nf1 regulates molecular functions in key biological signaling pathways via Ras. Nf1 affects diverse cellular functions by regulating several signaling pathways. Nf1 accelerates the conversion of active GTP-bound Ras into inactive GDP-bound Ras, thereby regulating numerous downstream effectors. Known signaling pathways downstream of Nf1 and Ras signaling include the Raf/MEK/ERK and PI3K/AKT/mTOR pathways. Figure created with BioRender.com and adapted from Anastasaki et al. (2022) [60], Masgras and Rasola (2021) [66], and Ratner and Miller (2015) [26]

can cause pain, disfigurement, and impair neurovascular structures and airways. Notably, pNFs are major contributors to the elevated mortality rates in NF1, as they can transform into malignant peripheral nerve sheath tumors (MPNST), which have a low survival rate [31, 88, 89]. Although surgical intervention is the standard treatment for pNF, it is typically palliative [88]. In 2020, selumetinib, a MEK inhibitor, gained United States Food and Drug Administration (FDA) approval for treating symptomatic and inoperable pNF in children [73–75].

In addition to neurofibromas, optic pathway gliomas (OPGs) are the second most common tumors in NF1—approximately 15–20% of children with NF1 develop OPGs [90]. Although OPGs are often non-lethal, about 30% of affected individuals will experience visual decline or loss due to these tumors, significantly reducing their quality of life. Given the severe delayed toxicity of radiotherapy and increased risk of visual loss with surgery, chemotherapy is the first line of treatment

for OPGs that cause visual decline. Notably, females are three times more likely to require treatment [90–95].

Experimental animal models of NF1 and in vitro cellular studies have provided significant insights into NF1-related tumor development. These models show significant alterations in growth, cell proliferation, and tumor progression. Such phenotypes stem from the interplay between multiple tissues, signaling pathways, neurite growth, and neuronal excitability. Introducing patient-derived NF1 mutations into hiPSCs impairs Schwann cell differentiation, promotes stemness, and fosters neurofibroma formation [96]. Furthermore, studies using hiPSCs and murine models have revealed that Nf1 mutations increase neuronal excitability, exacerbating tumor progression in both the central and peripheral nervous system [97–99]. Neuronal activity and midline expression directly impact the development and progression of mouse *Nf1*-OPG [97, 98]. Importantly, the progression of optic glioma growth in *Nf1* mutant mice can be selectively suppressed with clinically relevant dosing of lamotrigine, an anti-epileptic drug, for months after treatment cessation [100].

Aberrant regulation of the Raf/MEK/ERK signaling pathway plays a pivotal role of NF1-related tumorigenesis [26, 101], and MEK is a therapeutic target [73–75, 102, 103]. Both human and mouse models of MPNST exhibit aberrant activation of ERK (one molecular step downstream of MEK). Targeted pharmacological inhibition of the Raf/MEK/ERK pathway has been shown to inhibit tumor progression [101]. Proteomic analysis has also revealed Ras/PI3K-dependent activation of mTOR signaling in astrocytes from human or mutant mice optic nerve gliomas [63]. In NF1-deficient cells and human tumors, mTOR is constitutively activated [63, 64]. Notably, pharmacological inhibition of mTOR, MEK, and AKT signaling can restore normal proliferation in *Nf1*-deficient astrocytes [63, 104]. Additional pathways, such as cAMP/PKA, are also targets of interest for therapeutic interventions, playing roles in regulating cell differentiation and growth arrest [78]. In *Drosophila*, Nf1 regulates growth through non-cell-autonomous control of cAMP/PKA signaling in neuroendocrine cells [68]. In human neural progenitor cells, loss of Nf1 decreases cAMP levels, resulting in smaller growth cone areas and shorter axonal lengths [82]. These neural deficits can be restored through increased cAMP levels and by inhibiting Ras activity [82].

The zebrafish model, known for its transparent embryos, offers a unique lens through which to study the role of Nf1 during development. The zebrafish genome contains two *NF1* orthologs, *nf1a* and *nf1b*, each with over 90% similarity to human *NF1* at the amino acid level [105]. Experiments involving transient knockdown

of these *nf1* orthologs during embryogenesis result in vascular patterning defects, echoing observations seen in murine NF1 models and mirroring hallmarks of the human disease [105]. Furthermore, *nf1a* and *nf1b* zebrafish larvae exhibit hyperplasia of oligodendrocyte progenitor and Schwann cells [106, 107]. Additionally, *nf1* knockout initiates gliomagenesis in adult zebrafish brain tissue [108].

NF1 mutations introduced into Yucatan miniature pigs (minipigs) mimic characteristics commonly observed in NF1 patients. Two mutations have been introduced, which model prevalent human *NF1* mutations: *NF1*^{R1947X}, representing a common nonsense mutation, and *NF1*^{+/_{ex42del}}, emulating a heterozygous *NF1* mutation [109, 110]. Minipigs with either of these *NF1* mutations exhibit major clinical hallmarks of NF1, including café-au-lait macules (CALMs), OPGs, and neurofibromas [109, 110]. Notably, the minipig is unique among model organisms in that it exhibits spontaneous loss of NF1 heterozygosity, which drives tumor formation in humans [109].

Behavioral deficits and neuronal alterations

Cognitive impairment is a prevalent complication of NF1, affecting approximately 80% of those diagnosed with NF1 [22, 111]. Individuals with NF1 are significantly more likely to encounter a spectrum of developmental delays, such as deficits in learning, memory, executive function, broad language deficits, and fine motor skills [22, 29, 112]. NF1 patients may exhibit below-average IQ scores, with a small subset (4–8%) falling into the intellectually impaired range [22, 113]. The disorder is also highly comorbid with attention-deficit/hyperactivity disorder (ADHD) and ASD. Approximately half of the children with NF1 are diagnosed with ADHD [22, 111], and 12–49% exhibit symptoms of ASD [114–118]. These cognitive and behavioral challenges significantly impact quality of life of NF1 patients, affecting their emotional well-being, physical health, role functioning, and social interactions [25].

Given that NF1 increases risk for cognitive/behavioral symptoms, a major question is how loss of neurofibromin affects neuronal/brain function. Studies utilizing various animal models, including flies, zebrafish, mice, and minipigs, have contributed to understanding the role of Nf1 function in the nervous system. The *Drosophila* Nf1 protein, sharing 60% amino acid sequence homology with its human counterpart and conserved Ras GAP functionality [80], serves as an outstanding model for investigating genetics, neuronal function, and molecular signaling pathways in vivo [119–121]. In *Drosophila*, Nf1 is ubiquitously expressed during development and is prominently localized in the adult nervous system [39]. Loss of Nf1 function in flies disrupts sleep and circadian rhythms

[122–124]; the circadian rhythm deficit can be rescued by restoring the expression of wild-type *Drosophila* Nf1 in neurons or by attenuating Ras/ERK signaling pathways [122]. Additionally, *Drosophila* Nf1 mutants exhibit learning and memory deficits, including impaired olfactory associative learning and deficits in short-, middle-, and long-term memory [67, 79, 125, 126]. These learning and memory impairments can be rescued by restoring wild-type Nf1 protein in a neuron-specific manner [125] and ameliorated by enhancing PKA activity [79, 80]. Additionally, pharmacologically and genetically attenuating ALK, an upstream RTK, rescues associative learning deficits in *nf1* mutants [67].

Mutations in the *Drosophila* *NF1* ortholog increase locomotor activity and spontaneous grooming [127, 128], phenotypic analogs of the ADHD symptoms common in NF1 patients [29, 111]. Nf1/Ras signaling regulates grooming behavior, as the Nf1 GRD is required in neurons to maintain normal levels of grooming in *Drosophila* [127]. Besides motor-related behaviors, *Drosophila* *nf1* mutants display social and behavioral alterations, including delayed flight and climbing responses and altered sleep patterns [80, 122, 129]. Loss of Nf1 alters social behavior, specifically male courtship [130]. Synaptic transmission at the neuromuscular junction is altered in *nf1* mutants, suggesting that synaptic physiology changes may contribute to the phenotypes [131, 132].

Murine models of NF1 have been invaluable in unraveling Nf1's functions within the nervous system via structural plasticity and modulation of signaling pathways. In the rat hippocampus, the loss of Nf1 function disrupts pyramidal dendritic spine structural plasticity, resulting in the activity-dependent loss of dendritic spines due to sustained Ras activation [133]. In mice, Nf1 haploinsufficiency (*Nf1*[±]) replicate cognitive and behavioral deficits observed in NF1 patients, manifesting as deficits in hippocampal spatial learning and reduced long-term potentiation driven by increased GABA-mediated inhibition [44, 45, 48]. Rescue of Raf/MEK/ERK activity, either pharmacologically or genetically, ameliorates learning deficits and rescues long-term potentiation [44, 134, 135]. Moreover, *Nf1*[±] mice show heightened excitability in sensory neurons. Along with dopamine deficiency, this could contribute to learning impairment [99, 136–138]. Additionally, both human *NF1* and mouse *Nf1* are enriched in inhibitory neurons within the cortex [139]. Nf1 plays a crucial role in the nervous system beyond cognition and physiology, as human-derived *Nf1* mutations increase neuronal excitability in mice, accelerating tumor progression in the central and peripheral nervous system [97, 98].

Several other vertebrate models like zebrafish and minipigs have recapitulated neurocognitive deficits similar to

those observed in NF1 patients. Zebrafish with *nf1* mutations display learning and memory deficits, including short- and long-term habituation; these can be restored either through pharmacological inhibition of Ras downstream targets or by increasing cAMP signaling [81, 107]. The *NF1*^{+/*ex42del*} mutation in the minipig model produces neurocognitive deficits akin to those observed in NF1 patients, including learning and memory impairments and hyperactivity [110]. In addition, the NF1 minipig model exhibits altered pain sensitivity associated with NF1—examination of dorsal root ganglia expressing mutant *NF1*^{+/*ex42del*} revealed dysregulation of calcium and sodium channels [110]. Overall, these findings underscore the importance of Nf1 function in regulating neuronal development, structure, activity, and function.

Metabolic alterations

Metabolism is altered in multiple ways in NF1, and these changes may contribute to the pathophysiology of the disease. Patients with NF1 exhibit systemic metabolic shifts (Fig. 4) [32, 140]. Studies on body composition reveal multiple anomalies, including a lower body mass index (BMI) [141], reduced triglyceride stores [142], decreased bone mineral density [143], and shorter stature relative to unaffected individuals [144]. NF1 patients display lower muscle function [141], reduced maximal muscular strength [24, 145], and compromised motor proficiency [146]. In a comprehensive analysis of resting energy expenditure (REE), women with NF1 display heightened REE despite lower BMI [24]. Additionally, NF1 patients have a lower respiratory quotient (RQ), which indicates a differential reliance on fat oxidation over carbohydrate metabolism [24].

In addition to altered body composition, individuals with NF1 present a metabolic profile characterized by lower fasting blood glucose levels [147], heightened insulin sensitivity [148], and a reduced incidence of diabetes mellitus [140, 149, 150]. Hormonal dysregulation in NF1 patients, involving alterations in leptin, vistafin, and adiponectin, may contribute to these metabolic features [148]. Also noted are decreased levels of calcium, calcitonin, and vitamin D [143]. In adults with NF1, there is a notable reduction in cerebral glucose metabolism in the thalamus, as evidenced by positron emission tomography scans [23]. In addition to these differences, individuals with NF1 often experience significant cognitive and physiological fatigue [151], suggesting that metabolic dysregulation may impact brain function.

The Ras/Raf/MEK/ERK pathway regulates multiple metabolic processes, including cell proliferation, protein synthesis, lipid and cholesterol homeostasis, adipocyte differentiation, lipolysis, lipogenesis, gluconeogenesis, and gene expression (Fig. 3) [152, 153]. Consequently,

Metabolic-related symptoms of NF1 patients

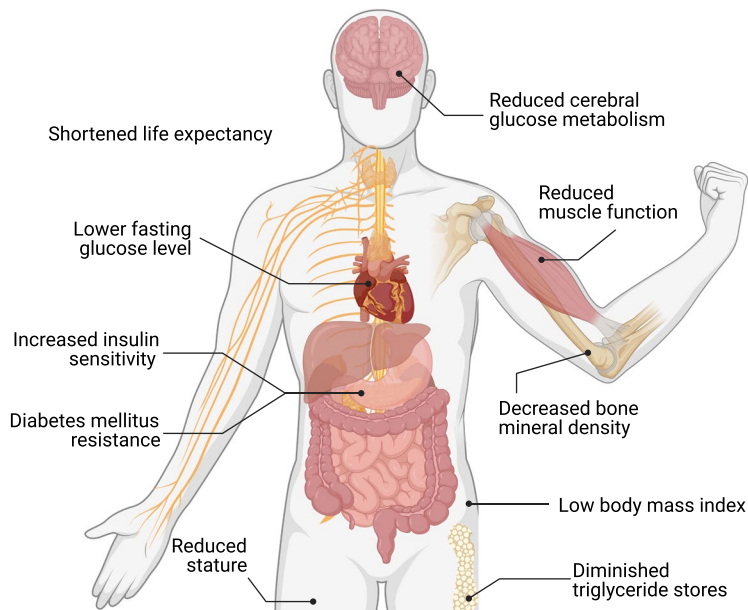


Fig. 4 Neurofibromatosis type 1 metabolic-related symptoms. Multisystemic alterations in metabolism that are commonly associated with NF1. Modified from Masgras and Rasola (2021) [66]. Created with BioRender.com






<i>In vitro</i> model	Murine model	<i>Drosophila</i> model	Zebrafish model	Minipig model
 Cellular bioenergetics Tumor development Neuronal activity Signaling pathway analysis	 Cognitive deficits (learning and memory) Metabolic alterations (tissue specific) Neuronal activity (developmental defects, dopamine deficiency, neuronal excitability)	 Metabolic alterations (whole-body and neuronal) Cognitive deficits (learning and memory) Activity alterations	 Developmental effects Cognitive deficits (habituation) Motor deficits	 Clinical hallmarks (neurofibromas, optic pathway gliomas, café-au-lait macules) Cognitive deficits

Fig. 5 Animal and in vitro models of NF1. Strengths of in vitro, murine, *Drosophila*, zebrafish, and minipig models to investigate NF1. Created with BioRender.com

hyperactivation of this pathway due to loss of *Nf1* may mediate the observed cellular and systemic metabolic dysfunctions. *Nf1*-deficient cells exhibit increased glycolysis and reduced mitochondrial respiration mediated through the Ras/MEK/ERK pathway [154].

Metabolic features of the disorder have been recapitulated in animal models of NF1 (Fig. 5). Heterozygous *Nf1* (*Nf1*[±]) mice exhibit altered body composition, represented by a reduction in fat mass and increased

percentage of lean mass [155]. Similar to NF1 patients, the loss of *Nf1* function enhances insulin sensitivity and glucose utilization in *Nf1*[±] mice [148, 155]. Conditional *Nf1* knockout results in metabolic changes in muscles, including reduced muscle growth, increased triglyceride content, malformations (cardiac, renal, hepatic, and skeletal muscle defects), and prenatal lethality [155–157]. Inactivation of *Nf1* in skeletal muscle (*Nf1*^{MyoD}^{-/-}) proves lethal within the first week of life; during development,

animals with the mutation exhibit stunted growth and intramyocellular lipid accumulation, indicative of impaired long chain fatty acid metabolism [156, 158]. Notably, muscle samples from limb-specific *Nf1* conditional knockout (*Nf1_{PxxI}^{-/-}*) mice recapitulate some of the pathological findings observed in human NF1 muscle biopsies, including intramyocellular lipid accumulation, elevated oxidative metabolic enzyme activity, heightened expression of leptin and fatty acid synthase, and reduced fatty acid transporters [156, 158].

Muscle weakness in NF1 may stem from changes in lipid storage resembling lipid storage myopathies. *Nf1* manipulations in mice suggest a role for *Nf1* in metabolic regulation within muscle tissue, suggesting avenues for potential therapeutic interventions. For example, when *Nf1* is lost in mesenchymal tissues (*Nf1_{PxxI}^{-/-}* mice), dietary interventions that reduce long chain fatty acid intake and enrich medium-fatty acids with L-carnitine effectively rescue lipid accumulation and muscle weakness [158]. Additionally, pharmacological intervention using the selective MEK inhibitor, PD98059, rescues postnatal body weight loss and lipid accumulation in mice with muscle-specific *Nf1* knockout when administered during pregnancy in *Nf1_{MyoD}^{-/-}* dams [159]. In pediatric NF1 patients, pharmacological inhibition of MEK with selumetinib or PD0325901 has led to clinically significant improvements in muscular strength [74], supporting a MEK/ERK-dependent mechanism underlying *Nf1*-associated muscle metabolism. Lastly, recent studies have highlighted the cell-autonomous role of *Nf1* in postnatal muscle growth and metabolic homeostasis, with homozygous *Nf1* mutations resulting in neonatal lethality [160]. Overall, these data suggest a critical role for *Nf1* in muscle development and function, providing insights into potential therapeutic interventions.

Research involving several models suggests that the metabolic alterations associated with NF1 extend beyond muscle tissue. In *Drosophila*, loss of *Nf1* drives multiple phenotypes indicative of metabolic dysfunction. *Nf1* mutations decrease body size by 15–25%, mirroring the short stature observed in NF1 patients [68, 80, 141, 161]. Restoring human or *Drosophila* *Nf1* expression in *nf1* mutant neurons can rescue the mutant growth defect [77, 161, 162]. Additionally, *Drosophila nf1* mutants experience a significant reduction in lifespan due to altered mitochondrial respiration and increased production of reactive oxygen species (ROS) [163]. Overexpressing *Nf1* in *nf1* mutants rescues lifespan, enhances mitochondrial respiration, and significantly reduces ROS production [163]. Furthermore, *Nf1* regulates metabolic homeostasis, with *Nf1* deficiency increasing metabolic rate (CO₂ production

and O₂ consumption), decreasing glycogen and triglyceride stores, and increasing the rate of lipid turnover [164, 165]. Similar to human NF1 patients, *Nf1* knockdown in *Drosophila* results in a reduced RQ [164], indicating increased fat utilization. These effects emanate, at least in part, from neuronal mechanisms [164]. Additionally, loss of *Nf1* heightens starvation susceptibility and increases feeding, likely in a compensatory manner to the metabolic alterations [164]. The metabolic phenotype in *Drosophila* resembles the increased REE observed in NF1 patients [24], suggesting metabolic dysfunction across different species (and cell types). Notably, the *Nf1* metabolic and motor (grooming) phenotypes are caused by the loss of *Nf1* function in different neural subsets, as knockdown in metabolism-regulating neurons does not affect grooming [164]. Collectively, these metabolic alterations highlight *Nf1*'s tight regulation of metabolic function and its susceptibility to disruption when *Nf1* is lost.

The molecular mechanism of *Nf1*'s effects on metabolism involves its activity as a Ras GAP. The catalytic activity within the GRD of *Nf1* is required for *Nf1*-dependent modulation of metabolic rate. A patient-derived mutation in the GRD (R1320P) fails to rescue metabolic phenotypes [164]. This mutation is at the equivalent residue as R1276P in humans, which reduces Ras-GAP activity by over 1000-fold without impairing any other *Nf1* function [61]. Transgenic expression of full-length, wild-type *Nf1* selectively in neurons of *nf1* mutants restores normal metabolic function [164] (as well as grooming activity) [127]. Additionally, the activation of downstream targets of *Nf1* and Ras, such as ERK, play a significant role in driving metabolic effects. Constitutive ERK activation in metabolism-regulating neurons increases metabolic rate, phenocopying the metabolic dysregulation observed in *Nf1* mutations [164]. Beyond neuronal and muscle-specific effects, metabolic profiling of mouse embryonic fibroblasts (MEFs) derived from *Nf1* knockout animals has revealed significant alterations in cellular bioenergetics. Specifically, *Nf1* knockout MEFs displayed diminished mitochondrial activity, driven by elevated glycolysis and decreased respiration [154]. These metabolic shifts stem from heightened Ras/MEK/ERK signaling within mitochondria [154]. Collectively, these results suggest that *Nf1* acts in neurons, muscles, and potentially additional cell types to regulate cellular bioenergetics in multiple model systems. Further, the mechanism involves the overactivation of Ras/Raf/MEK/ERK activity. The contributions of other downstream signaling pathways (e.g., the metabolism-regulating mTOR pathway) provide potentially promising avenues for exploration in the study of metabolic regulation in NF1.

Pigmentary lesions

Pigmentary features are crucial for early diagnosis of NF1. CALMs, which are observed in 99% of NF1 patients by age 1 [166], consist of melanocytes with biallelic *NFI* inactivation [10, 167]. These features are among the earliest signs of the disease. Additionally, axillary and inguinal freckling, typically appear between 3 to 5 years of age and are present in about 90% of patients by age 7 [166]. Another significant marker are Lisch nodules, which are asymptomatic hyperpigmented iris hamartomas, typically appear by age 5–6. These nodules are present in over 70% of patients by the age of 10 and are observed in over 90% of adults with NF1 [166, 168]. Two *NFI* mutations in minipig models, *NFI*^{R1947X} and *NFI*^{+/*ex42del*}, have successfully replicated these pigmentary features, including CALMs and axillary freckling [109, 110]. The loss of Nf1 expression in minipigs models of NF1 results in hyperactivation of the Ras pathway and its effector molecules, linking the signaling cascades to the pathogenesis of cutaneous NF1 features [110]. Finally, homozygous *nf1a* and *nf1b* mutant zebrafish larvae exhibit pigmentation anomalies, providing a novel vertebrate model to study pigmentation lesions associated with NF1 [107].

Skeletal abnormalities

Patients with NF1 exhibit a range of skeletal abnormalities, leading to significant morbidity. These osseous defects include both localized and generalized bone deformities, contributing to bone weakening and an increased fracture risk. One of the most significant manifestations is long-bone dysplasia, which affects approximately 5% of individuals with NF1. This condition is characterized by anterolateral bowing of the lower limbs, predominately affecting the tibia, resulting in decreased bone density, increased fracture risk, and pseudarthrosis [11, 92, 169, 170]. Another notable skeletal anomaly is sphenoid-wing dysplasia, which affects up to 11% of NF1 patients and results in distinct cranial deformities [88, 171, 172]. Scoliosis is the most prevalent skeletal defect associated with NF1, occurring in up to 30% of patients and often necessitating surgical intervention in severe cases [173]. Furthermore, NF1 patients tend to be shorter than their healthy counterparts, with 8–15% experiencing a generalized reduction in skeletal bone growth [174, 175]. Individuals with NF1 exhibit both local and general dysregulation of bone resorption and remodeling, leading to increased formation of osteoclast [176, 177]. NF1 patients often have a reduced bone mineral density, osteoporosis, and increased risk of bone fractures [178].

In animal models, Nf1 is critical in skeletal development. In mice, Nf1 is essential for joint development; conditional Nf1 loss during early limb development induces multiple joint abnormalities, including deformities in

the hip, knee, and elbow [179]. Similar to human NF1 patients, tibia bowing occurs in mice due to Nf1 deficiency, leading to growth retardation and abnormal growth plate development [179]. Moreover, the expression of Nf1 in bone marrow osteoprogenitors is crucial for maintaining adult skeletal integrity [180]. Loss of Nf1 in these cells leads to skeletal anomalies resembling those seen in NF1 patients, including progressive scoliosis, kyphosis, tibial bowing, and deformities in the skull and anterior chest wall [181]. Additionally, Nf1 loss in osteochondroprogenitors results in decreased bone mass, increased cortical porosity, severe short stature, and intervertebral disc defects [181]. Similar skeletal phenotypes are observed in a minipig model of NF1. The *NFI*^{+/*ex42del*} minipig model develops tibial bone curvature and shorter long bones such as the femur, tibia, humerus, ulna, metacarpals, indicative of reduced stature [182]. These animal models further substantiate the critical role of Nf1 in skeletal integrity and development.

Developmental alterations

The loss of Nf1 results in developmental alterations that may contribute to the clinical manifestations of NF1. Magnetic resonance imaging studies have documented alterations in neuronal development among NF1 patients, including increased total brain and white matter volumes. Notably, enlargements in subregions including corpus callosum and brainstem, as well as increased optic nerve tortuosity, are commonly observed [20, 183–196]. Along with structural differences, loss of Nf1 is linked to a range of functional changes in neuronal activity. These include changes in cortical association networks and functional connectivity within the default network, corticostriatal functional circuits, and areas critical for cognition, social functioning, executive functioning, and spatial working memory [197–205]. Collectively, these observations underscore the role of Nf1 in modulating brain development, connectivity, and function.

Conditional knockout of neuronal Nf1 in mice mirrors human pathology by enlarging the corpus callosum, an effect which can be rescued by inhibiting Raf/MEK/ERK signaling during neonatal development [206]. In a more severe manifestation, homozygous *Nf1* knockout (*Nf1*^{-/-}) mice exhibit gestational lethality due to severe cardiovascular abnormalities, highlighting a significant role for Nf1 during tissue development [157]. Heterozygous *Nf1* mutant (*Nf1*[±]) mice, although viable, exhibit numerous brain abnormalities, including enlarged glia, increased neuron numbers, astrocyte proliferation, and neural tube closure defects [157, 207–209]. Neuron-specific *Nf1* knockout in mice also reveals brain abnormalities, including abnormal cortex development, increased cell density, heightened astrocyte proliferation, and reduced

cortical thickness [210]. Loss of Nf1 function in neurons, rather than glia, in mice causes growth defects, further underscoring Nf1's critical role in neuronal growth and development [210, 211].

The structural alterations resulting from the loss of Nf1 raise significant questions about whether and how these developmental alterations influence behavior. Understanding this relationship is crucial for optimizing the timing of therapeutic interventions by, for instance, allowing for targeted treatments during appropriate developmental times. While direct behavioral correlations in humans are yet to be established, animal models have provided valuable insights. For example, in *Drosophila*, the developmental contribution of Nf1 to adult behavior has been parsed. Loss of Nf1 increases the frequency of spontaneous grooming behavior in adult animals [128]. Additionally, studies using conditional knockdown of Nf1 in neurons across developmental time windows revealed that loss of Nf1 during a critical developmental period impairs motor (grooming) behavior, whereas similar alterations either earlier (embryonic stage) or later (adult stage) do not have the same effect [127]. The mechanisms by which Nf1 loss impacts neuronal development in NF1 are diverse and complex. They may include altered cell growth, division, differentiation/specification, apoptosis, dendrite & axon targeting, synaptogenesis, activity-dependent synaptic refinement, hormone responsivity, and nutrient responsivity [210–213]. Future mechanistic studies are necessary to dissect how developmental disruptions due to the loss of Nf1 result in adult phenotypes.

Conclusions

Research utilizing animal models and in vitro studies has elucidated the significant effects of Nf1 in the nervous system and behavior, identifying its significance in normal development and function. Nf1 influences cellular and systemic physiology via multiple molecular and cellular mechanisms, including alterations in metabolism. Several major models, such as mice, *Drosophila*, minipigs, and zebrafish, have considerably advanced our understanding of Nf1's mechanistic role within the nervous system and its effects on metabolic regulation. Continued advancements in these areas hold promise for the development of novel targeted therapies and interventions aimed at improving the outcomes and quality of life for individuals with NF1.

Abbreviations

ADHD	Attention-deficit/hyperactivity disorder
ALK	Anaplastic Lymphoma Kinase
ASD	Autism spectrum disorder
BMI	Body mass index
CALM	Café-au-lait macule
cAMP	Cyclic adenosine monophosphate

CN	Cutaneous neurofibromas
CNS	Central nervous system
FDA	Food and Drug Administration
GAP	GTPase activating protein
GPCR	G protein-coupled receptor
GRD	GAP-related domain
hiPSC	Human induced pluripotent stem cells
MAPK	Mitogen-activated protein kinase
MEF	Mouse embryonic fibroblasts
MPNST	Malignant peripheral nerve sheath tumors
NF1	Neurofibromatosis type 1
<i>NF1</i>	Human neurofibromin 1 gene
<i>Nf1</i>	Non-human neurofibromin 1 gene
Nf1	Neurofibromin protein
<i>nf1</i>	Invertebrate mutant neurofibromin protein
OPG	Optic pathway glioma
PKA	Protein kinase A
PKC ζ	Protein kinase C zeta
pNF	Plexiform neurofibroma
REE	Resting energy expenditure
ROS	Reactive oxygen species
RQ	Respiratory quotient
RTK	Receptor tyrosine kinase

Acknowledgements

We thank Drs. Damon Page, Ronald L. Davis, Baoji Xu, and David Gutmann for advice and feedback during Dr. Botero's dissertation research, and Lisa Ringen, Linda Buckner, Melissa Benilous, and Katherine O'Brien for administrative support.

Authors' contributions

V.B. and S.M.T. conceived, wrote, and edited the manuscript. The authors read and approved the final manuscript.

Funding

Our research is supported by NIH/NINDS R01 NS097237, R01 NS126361, R01 NS114403, R01 NS124716, and R21 NS124198. V.B. was supported by NIH/NINDS F31 NS124245.

Availability of data and materials

N/A (Review).

Declarations

Ethics approval and consent to participate

Not applicable.

Consent of publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Received: 30 September 2023 Accepted: 2 August 2024

Published online: 31 August 2024

References

- Hirbe AC, Gutmann DH. Neurofibromatosis type 1: a multidisciplinary approach to care. *Lancet Neurol*. 2014;13(8):834–43.
- Uusitalo E, Leppävirta J, Koffert A, Suominen S, Vahtera J, Vahlberg T, et al. Incidence and mortality of neurofibromatosis: a total population study in finland. *J Invest Dermatol*. 2015;135(3):904–6.
- Evans DG, Howard E, Giblin C, Clancy T, Spencer H, Huson SM, et al. Birth incidence and prevalence of tumor-prone syndromes: estimates from a UK family genetic register service. *Am J Med Genet A*. 2010;152A(2):327–32.

4. Huson SM, Compston DA, Clark P, Harper PS. A genetic study of von Recklinghausen neurofibromatosis in south east Wales. I. Prevalence, fitness, mutation rate, and effect of parental transmission on severity. *J Med Genet.* 1989;26(11):704–11.
5. Sanchez GM, Siuda T. Ebers Papyrus case #873: a probable case of neurofibromatosis 1. *S D J Med.* 2002;55(12):529–35.
6. Ruggieri M, Praticò AD, Caltabiano R, Polizzi A. Early history of the different forms of neurofibromatosis from ancient Egypt to the British Empire and beyond: First descriptions, medical curiosities, misconceptions, landmarks, and the persons behind the syndromes. *Am J Med Genet A.* 2018;176(3):515–50.
7. Von Recklinghausen FD. *Über ide multiplen Fibrome der Haut und ihre beziehung zu den multiplen Neuromen.* Berlin: Hirschwald; 1882. p. 3–18.
8. Crump T. Advances in neurology: Translation of case reports in Ueber die multiplen Fibrome der Haut und ihre Beziehung zu den multiplen Neuromen by F. v. Recklinghausen. *Arch Neurol.* 1981;29:259–75.
9. Schull WJ, Neel JV. A clinical, pathological, and genetic study of multiple neurofibromatosis: Charles C. Thomas. 1956.
10. Neurofibromatosis: Conference Statement. *Arch Neurol.* 1988;45(5):575–8. <https://doi.org/10.1001/archneur.1988.00520290115023>.
11. Legius E, Messiaen L, Wolkenstein P, Pancza P, Avery RA, Berman Y, et al. Revised diagnostic criteria for neurofibromatosis type 1 and Legius syndrome: an international consensus recommendation. *Genet Med.* 2021;23(8):1506–13.
12. Barker D, Wright E, Nguyen K, Cannon L, Fain P, Goldgar D, et al. Gene for von Recklinghausen neurofibromatosis is in the pericentromeric region of chromosome 17. *Science.* 1987;236(4805):1100–2.
13. Seizinger BR, Rouleau GA, Ozelius LJ, Lane AH, Faryniarz AG, Chao MV, et al. Genetic linkage of von Recklinghausen neurofibromatosis to the nerve growth factor receptor gene. *Cell.* 1987;49(5):589–94.
14. Collins FS, O'Connell P, Ponder BAJ, Seizinger BR. Progress towards identifying the neurofibromatosis (NF1) gene. *Trends Genet.* 1989;5:217–21.
15. Ballester R, Marchuk D, Boguski M, Saulino A, Letcher R, Wigler M, et al. The NF1 locus encodes a protein functionally related to mammalian GAP and yeast IRA proteins. *Cell.* 1990;63(4):851–9.
16. Martin GA, Viskochil D, Bollag G, McCabe PC, Crosier WJ, Haubruck H, et al. The GAP-related domain of the neurofibromatosis type 1 gene product interacts with ras p21. *Cell.* 1990;63(4):843–9.
17. Wallace MR, Marchuk DA, Andersen LB, Letcher R, Odeh HM, Saulino AM, et al. Type 1 neurofibromatosis gene: identification of a large transcript disrupted in three NF1 patients. *Science.* 1990;249(4965):181–6.
18. DeClue JE, Cohen BD, Lowy DR. Identification and characterization of the neurofibromatosis type 1 protein product. *Proc Natl Acad Sci U S A.* 1991;88(22):9914–8.
19. Gutmann DH, Wood DL, Collins FS. Identification of the neurofibromatosis type 1 gene product. *Proc Natl Acad Sci.* 1991;88(21):9658–62.
20. Pride N, Payne JM, Webster R, Shores EA, Rae C, North KN. Corpus callosum morphology and its relationship to cognitive function in neurofibromatosis type 1. *J Child Neurol.* 2010;25(7):834–41.
21. Gill DS, Hyman SL, Steinberg A, North KN. Age-related findings on MRI in neurofibromatosis type 1. *Pediatr Radiol.* 2006;36(10):1048–56.
22. Hyman SL, Shores A, North KN. The nature and frequency of cognitive deficits in children with neurofibromatosis type 1. *Neurology.* 2005;65(7):1037–44.
23. Apostolova I, Derlin T, Salamon J, Amthauer H, Granstrom S, Brenner W, et al. Cerebral glucose metabolism in adults with neurofibromatosis type 1. *Brain Res.* 2015;1625:97–101.
24. Souza MLR, Jansen AK, Rodrigues LOC, Vilela DLS, Kakehasi AM, Martins AS, et al. Increased resting metabolism in neurofibromatosis type 1. *Clin Nutr ESPEN.* 2019;32:44–9.
25. Gutmann DH, Ferner RE, Listerick RH, Korf BR, Wolters PL, Johnson KJ. Neurofibromatosis type 1. *Nat Rev Dis Primers.* 2017;3:17004.
26. Ratner N, Miller SJ. A RASopathy gene commonly mutated in cancer: the neurofibromatosis type 1 tumour suppressor. *Nat Rev Cancer.* 2015;15(5):290–301.
27. Upadhyaya M, Cooper DN. *Neurofibromatosis Type 1.* Heidelberg: Springer; 2012.
28. Campian J, Gutmann DH. CNS Tumors in Neurofibromatosis. *J Clin Oncol.* 2017;35(21):2378–85.
29. Hyman SL, Arthur E, North KN. Learning disabilities in children with neurofibromatosis type 1: subtypes, cognitive profile, and attention-deficit-hyperactivity disorder. *Dev Med Child Neurol.* 2006;48(12):973–7.
30. Stewart DR, Korf BR, Nathanson KL, Stevenson DA, Yohay K. Care of adults with neurofibromatosis type 1: a clinical practice resource of the American College of Medical Genetics and Genomics (ACMG). *Genet Med.* 2018;20(7):671–82.
31. Evans DGR, O'Hara C, Wilding A, Ingham SL, Howard E, Dawson J, et al. Mortality in neurofibromatosis 1: in North West England: an assessment of actuarial survival in a region of the UK since 1989. *Eur J Hum Genet.* 2011;19(11):1187–91.
32. Rasmussen SA, Yang Q, Friedman JM. Mortality in neurofibromatosis 1: an analysis using U.S. death certificates. *Am J Hum Genet.* 2001;68(5):1110–8.
33. Viskochil D, Buchberg AM, Xu G, Cawthon RM, Stevens J, Wolff RK, et al. Deletions and a translocation interrupt a cloned gene at the neurofibromatosis type 1 locus. *Cell.* 1990;62(1):187–92.
34. Shen MH, Harper PS, Upadhyaya M. Molecular genetics of neurofibromatosis type 1 (NF1). *J Med Genet.* 1996;33(1):2–17.
35. Li C, Cheng Y, Gutmann DA, Mangoura D. Differential localization of the neurofibromatosis 1 (NF1) gene product, neurofibromin, with the F-actin or microtubule cytoskeleton during differentiation of telencephalic neurons. *Dev Brain Res.* 2001;130(2):231–48.
36. Stowe IB, Mercado EL, Stowe TR, Bell EL, Oses-Prieto JA, Hernandez H, et al. A shared molecular mechanism underlies the human rasopathies Legius syndrome and Neurofibromatosis-1. *Genes Dev.* 2012;26(13):1421–6.
37. Nordlund M, Gu X, Shipley M, Ratner N. Neurofibromin is enriched in the endoplasmic reticulum of CNS neurons. *J Neurosci.* 1993;13(4):1588–600.
38. Roudebush M, Slabe T, Sundaram V, Hoppel CL, Golubic M, Stacey DW. Neurofibromin Colocalizes with Mitochondria in Cultured Cells. *Exp Cell Res.* 1997;236(1):161–72.
39. Daston MM, Scrabble H, Nordlund M, Sturbaum AK, Nissen LM, Ratner N. The protein product of the neurofibromatosis type 1 gene is expressed at highest abundance in neurons, Schwann cells, and oligodendrocytes. *Neuron.* 1992;8(3):415–28.
40. Philpott C, Tovell H, Frayling IM, Cooper DN, Upadhyaya M. The NF1 somatic mutational landscape in sporadic human cancers. *Hum Genomics.* 2017;11(1):13.
41. Easton DF, Ponder MA, Huson SM, Ponder BA. An analysis of variation in expression of neurofibromatosis (NF) type 1 (NF1): evidence for modifying genes. *Am J Hum Genet.* 1993;53(2):305–13.
42. Anastasaki C, Woo AS, Messiaen LM, Gutmann DH. Elucidating the impact of neurofibromatosis-1 germline mutations on neurofibromin function and dopamine-based learning. *Hum Mol Genet.* 2015;24(12):3518–28.
43. Bajenaru ML, Donahoe J, Corral T, Reilly KM, Brophy S, Pellicer A, et al. Neurofibromatosis 1 (NF1) heterozygosity results in a cell-autonomous growth advantage for astrocytes. *Glia.* 2001;33(4):314–23.
44. Costa RM, Federov NB, Kogan JH, Murphy GG, Stern J, Ohno M, et al. Mechanism for the learning deficits in a mouse model of neurofibromatosis type 1. *Nature.* 2002;415(6871):526–30.
45. Cui Y, Costa RM, Murphy GG, Elgersma Y, Zhu Y, Gutmann DH, et al. Neurofibromin regulation of ERK signaling modulates GABA release and learning. *Cell.* 2008;135(3):549–60.
46. Geist RT, Gutmann DH. Expression of a developmentally-regulated neuron-specific isoform of the neurofibromatosis 1 (NF1) gene. *Neurosci Lett.* 1996;211(2):85–8.
47. Gutmann DH, Andersen LB, Cole JL, Swaroop M, Collins FS. An alternatively-spliced mRNA in the carboxy terminus of the neurofibromatosis type 1 (NF1) gene is expressed in muscle. *Hum Mol Genet.* 1993;2(7):989–92.
48. Costa RM, Yang T, Huynh DP, Pulst SM, Viskochil DH, Silva AJ, et al. Learning deficits, but normal development and tumor predisposition, in mice lacking exon 23a of Nf1. *Nat Genet.* 2001;27(4):399–405.
49. Gutmann DH, Cole JL, Collins FS. Expression of the neurofibromatosis type 1 (NF1) gene during mouse embryonic development. *Prog Brain Res.* 1995;105:327–35.

50. Xu H, Gutmann DH. Mutations in the GAP-related domain impair the ability of neurofibromin to associate with microtubules. *Brain Res.* 1997;759(1):149–52.
51. Gutmann DH, Geist RT, Rose K, Wright DE. Expression of two new protein isoforms of the neurofibromatosis type 1 gene product, neurofibromin, in muscle tissues. *Dev Dyn.* 1995;202(3):302–11.
52. Gutmann DH, Zhang Y, Hirbe A. Developmental regulation of a neuron-specific neurofibromatosis 1 isoform. *Ann Neurol.* 1999;46(5):777–82.
53. Summers MA, Quinlan KG, Payne JM, Little DG, North KN, Schindeler A. Skeletal muscle and motor deficits in Neurofibromatosis Type 1. *J Musculoskelet Neuronal Interact.* 2015;15(2):161–70.
54. Gutman DH, Cole JL, Collins FS. Modulation of neurofibromatosis type 1 gene expression during in vitro myoblast differentiation. *J Neurosci Res.* 1994;37(3):398–405.
55. Kaufmann D, Müller R, Kenner O, Leistner W, Hein C, Vogel W, et al. The N-terminal splice product NF1-10a-2 of the NF1 gene codes for a transmembrane segment. *Biochem Biophys Res Commun.* 2002;294(2):496–503.
56. Gutmann DH, Geist RT, Wright DE, Snider WD. Expression of the neurofibromatosis 1 (NF1) isoforms in developing and adult rat tissues. *Cell Growth Differ.* 1995;6(3):315–23.
57. Vandenbroucke I, Vandesompele J, De Paepe A, Messiaen L. Quantification of NF1 transcripts reveals novel highly expressed splice variants. *FEBS Lett.* 2002;522(1–3):71–6.
58. Young LC, de GoldsteinSalazar R, Han SW, Huang ZYS, Merk A, Drew M, et al. Destabilizing NF1 variants act in a dominant negative manner through neurofibromin dimerization. *Proc Natl Acad Sci U S A.* 2023;120(5):e2208960120.
59. Mo J, Moye SL, McKay RM, Le LQ. Neurofibromin and suppression of tumorigenesis: beyond the GAP. *Oncogene.* 2022;41:1235–51.
60. Anastasaki C, Orozco P, Gutmann DH. RAS and beyond: the many faces of the neurofibromatosis type 1 protein. *Dis Models Mech.* 2022;15(2):dmm049362.
61. Klose A, Ahmadian MR, Schuelke M, Scheffzek K, Hoffmeyer S, Gewies A, et al. Selective disactivation of neurofibromin GAP activity in neurofibromatosis type 1. *Hum Mol Genet.* 1998;7(8):1261–8.
62. Scheffzek K, Ahmadian MR, Wiesmüller L, Kabsch W, Stege P, Schmitz F, et al. Structural analysis of the GAP-related domain from neurofibromin and its implications. *Embo j.* 1998;17(15):4313–27.
63. Dasgupta B, Yi Y, Chen DY, Weber JD, Gutmann DH. Proteomic analysis reveals hyperactivation of the mammalian target of rapamycin pathway in neurofibromatosis 1-associated human and mouse brain tumors. *Cancer Res.* 2005;65(7):2755–60.
64. Johannessen CM, Reczek EE, James MF, Brems H, Legius E, Cichowski K. The NF1 tumor suppressor critically regulates TSC2 and mTOR. *Proc Natl Acad Sci U S A.* 2005;102(24):8573–8.
65. Scheffzek K, Shivalingaiah G. Ras-specific GTPase-activating proteins: structures, mechanisms, and interactions. *Cold Spring Harb Perspect Med.* 2019;9(3):a031500.
66. Masgras I, Rasola A. Metabolic Features of Neurofibromatosis Type 1-Associated Tumors. In: Juichiro N, Yuichi Y, editors. *Clinical and Basic Aspects of Neurofibromatosis Type 1.* Rijeka: IntechOpen; 2021. p. Ch. 9.
67. Gouzi JY, Moressis A, Walker JA, Apostolopoulou AA, Palmer RH, Bernardis A, et al. The receptor tyrosine kinase Alk controls neurofibromin functions in *Drosophila* growth and learning. *PLoS Genet.* 2011;7(9):e1002281.
68. Walker JA, Gouzi JY, Long JB, Huang S, Maher RC, Xia H, et al. Genetic and functional studies implicate synaptic overgrowth and ring gland cAMP/PKA signaling defects in the *Drosophila melanogaster* neurofibromatosis-1 growth deficiency. *PLoS Genet.* 2013;9(11):e1003958.
69. Lorén CE, Scully A, Grabbe C, Edeen PT, Thomas J, McKeown M, et al. Identification and characterization of DAik: a novel *Drosophila melanogaster* RTK which drives ERK activation in vivo. *Genes Cells.* 2001;6(6):531–44.
70. Palmer RH, Vernersson E, Grabbe C, Hallberg B. Anaplastic lymphoma kinase: signalling in development and disease. *Biochem J.* 2009;420(3):345–61.
71. Xie K, Colgan LA, Dao MT, Muntean BS, Sutton LP, Orlandi C, et al. NF1 is a direct G protein effector essential for opioid signaling to Ras in the striatum. *Current biology : CB.* 2016;26(22):2992–3003.
72. Sánchez-Alegria K, Flores-León M, Avila-Muñoz E, Rodríguez-Corona N, Arias C. PI3K signaling in neurons: a central node for the control of multiple functions. *Int J Mol Sci.* 2018;19(12):3725.
73. Mukhopadhyay S, Maitra A, Choudhury S. Selumetinib: the first ever approved drug for neurofibromatosis-1 related inoperable plexiform neurofibroma. *Curr Med Res Opin.* 2021;37(5):789–94.
74. Gross AM, Wolters PL, Dombi E, Baldwin A, Whitcomb P, Fisher MJ, et al. Selumetinib in Children with Inoperable Plexiform Neurofibromas. *N Engl J Med.* 2020;382(15):1430–42.
75. Bergqvist C, Wolkenstein P. MEK inhibitors in RASopathies. *Curr Opin Oncol.* 2021;33(2):110–9.
76. Dasgupta B, Dugan LL, Gutmann DH. The neurofibromatosis 1 gene product neurofibromin regulates pituitary adenylate cyclase-activating polypeptide-mediated signaling in astrocytes. *J Neurosci.* 2003;23(26):8949–54.
77. Tong J, Hannan F, Zhu Y, Bernardis A, Zhong Y. Neurofibromin regulates G protein-stimulated adenylyl cyclase activity. *Nat Neurosci.* 2002;5(2):95–6.
78. Kim HA, Ratner N, Roberts TM, Stiles CD. Schwann cell proliferative responses to cAMP and NF1 are mediated by cyclin D1. *J Neurosci.* 2001;21(4):1110–6.
79. Guo HF, Tong J, Hannan F, Luo L, Zhong Y. A neurofibromatosis-1-regulated pathway is required for learning in *Drosophila*. *Nature.* 2000;403(6772):895–8.
80. The I, Hannigan GE, Cowley GS, Reginald S, Zhong Y, Gusella JF, et al. Rescue of a *Drosophila* NF1 mutant phenotype by protein kinase A. *Science.* 1997;276(5313):791–4.
81. Wolman MA, de Groh ED, McBride SM, Jongens TA, Granato M, Epstein JA. Modulation of cAMP and Ras signaling pathways improves distinct behavioral deficits in a zebrafish model of neurofibromatosis type 1. *Cell Rep.* 2014;8(5):1265–70.
82. Anastasaki C, Gutmann DH. Neuronal NF1/RAS regulation of cyclic AMP requires atypical PKC activation. *Hum Mol Genet.* 2014;23(25):6712–21.
83. Brown JA, Diggs-Andrews KA, Gianino SM, Gutmann DH. Neurofibromatosis-1 heterozygosity impairs CNS neuronal morphology in a cAMP/PKA/ROCK-dependent manner. *Mol Cell Neurosci.* 2012;49(1):13–22.
84. Jouhilahti EM, Peltonen S, Heape AM, Peltonen J. The pathoetiology of neurofibromatosis 1. *Am J Pathol.* 2011;178(5):1932–9.
85. Laycock-van Spyk S, Thomas N, Cooper DN, Upadhyaya M. Neurofibromatosis type 1-associated tumours: their somatic mutational spectrum and pathogenesis. *Hum Genomics.* 2011;5(6):623–90.
86. Dombi E, Solomon J, Gillespie AJ, Fox E, Balis FM, Patronas N, et al. NF1 plexiform neurofibroma growth rate by volumetric MRI: relationship to age and body weight. *Neurology.* 2007;68(9):643–7.
87. Reed N, Gutmann DH. Tumorigenesis in neurofibromatosis: new insights and potential therapies. *Trends Mol Med.* 2001;7(4):157–62.
88. Choi J, An S, Lim SY. Current concepts of neurofibromatosis type 1: pathophysiology and treatment. *Arch Craniofac Surg.* 2022;23(1):6–16.
89. Hwang IK, Hahn SM, Kim HS, Kim SK, Kim HS, Shin KH, et al. Outcomes of treatment for malignant peripheral nerve sheath tumors: different clinical features associated with neurofibromatosis Type 1. *Cancer Res Treat.* 2017;49(3):717–26.
90. Listernick R, Charrow J, Greenwald M, Mets M. Natural history of optic pathway tumors in children with neurofibromatosis type 1: a longitudinal study. *J Pediatr.* 1994;125(1):63–6.
91. Ferner RE. Neurofibromatosis 1 and neurofibromatosis 2: a twenty first century perspective. *Lancet Neurol.* 2007;6(4):340–51.
92. Cimino PJ, Gutmann DH. Chapter 51 - Neurofibromatosis type 1. In: Geschwind DH, Paulson HL, Klein C, editors. *Handbook of Clinical Neurology.* 148. Amsterdam: Elsevier; 2018. p. 799–811.
93. Diggs-Andrews KA, Brown JA, Gianino SM, Rubin JB, Wozniak DF, Gutmann DH. Sex is a major determinant of neuronal dysfunction in neurofibromatosis type 1. *Ann Neurol.* 2014;75(2):309–16.
94. Fisher MJ, Loguidice M, Gutmann DH, Listernick R, Ferner RE, Ullrich NJ, et al. Visual outcomes in children with neurofibromatosis type 1-associated optic pathway glioma following chemotherapy: a multicenter retrospective analysis. *Neuro Oncol.* 2012;14(6):790–7.
95. Sani I, Albanese A. Endocrine long-term follow-up of children with neurofibromatosis type 1 and optic pathway glioma. *Horm Res Paediatr.* 2017;87(3):179–88.

96. Mo J, Anastasaki C, Chen Z, Shipman T, Papke J, Yin K, et al. Humanized neurofibroma model from induced pluripotent stem cells delineates tumor pathogenesis and developmental origins. *J Clin Investig.* 2021;131(1):e139807.
97. Anastasaki C, Mo J, Chen JK, Chatterjee J, Pan Y, Scheaffer SM, et al. Neuronal hyperexcitability drives central and peripheral nervous system tumor progression in models of neurofibromatosis-1. *Nat Commun.* 2022;13(1):2785.
98. Pan Y, Hysinger JD, Barron T, Schindler NF, Cobb O, Guo X, et al. NF1 mutation drives neuronal activity-dependent initiation of optic glioma. *Nature.* 2021;594(7862):277–82.
99. Wang Y, Nicol GD, Clapp DW, Hingtgen CM. Sensory neurons from NF1 haploinsufficient mice exhibit increased excitability. *J Neurophysiol.* 2005;94(6):3670–6.
100. Anastasaki C, Chatterjee J, Koleske JP, Gao Y, Bozeman SL, Kernan CM, et al. NF1 mutation-driven neuronal hyperexcitability sets a threshold for tumorigenesis and therapeutic targeting of murine optic glioma. *Neuro Oncol.* 2024;26:1496–508.
101. Jessen WJ, Miller SJ, Jousma E, Wu J, Rizvi TA, Brundage ME, et al. MEK inhibition exhibits efficacy in human and mouse neurofibromatosis tumors. *J Clin Investig.* 2013;123(1):340–7.
102. Dombi E, Baldwin A, Marcus LJ, Fisher MJ, Weiss B, Kim A, et al. Activity of Selumetinib in Neurofibromatosis Type 1-Related Plexiform Neurofibromas. *N Engl J Med.* 2016;375(26):2550–60.
103. Gross AM, Dombi E, Widemann BC. Current status of MEK inhibitors in the treatment of plexiform neurofibromas. *Childs Nerv Syst.* 2020;36(10):2443–52.
104. Kaul A, Toonen JA, Cimino PJ, Gianino SM, Gutmann DH. Akt- or MEK-mediated mTOR inhibition suppresses Nf1 optic glioma growth. *Neuro Oncol.* 2015;17(6):843–53.
105. Padmanabhan A, Lee JS, Ismat FA, Lu MM, Lawson ND, Kanki JP, et al. Cardiac and vascular functions of the zebrafish orthologues of the type I neurofibromatosis gene NFI. *Proc Natl Acad Sci U S A.* 2009;106(52):22305–10.
106. Lee JS, Padmanabhan A, Shin J, Zhu S, Guo F, Kanki JP, et al. Oligodendrocyte progenitor cell numbers and migration are regulated by the zebrafish orthologs of the NF1 tumor suppressor gene. *Hum Mol Genet.* 2010;19(23):4643–53.
107. Shin J, Padmanabhan A, de Groh ED, Lee JS, Haidar S, Dahlberg S, et al. Zebrafish neurofibromatosis type 1 genes have redundant functions in tumorigenesis and embryonic development. *Dis Model Mech.* 2012;5(6):881–94.
108. Luo J, Liu P, Lu C, Bian W, Su D, Zhu C, et al. Stepwise crosstalk between aberrant Nf1, Tp53 and Rb signalling pathways induces gliomagenesis in zebrafish. *Brain.* 2021;144(2):615–35.
109. Isakson SH, Rizzardi AE, Coutts AW, Carlson DF, Kirstein MN, Fisher J, et al. Genetically engineered minipigs model the major clinical features of human neurofibromatosis type 1. *Commun Biol.* 2018;1(1):158.
110. White KA, Swier VJ, Cain JT, Kohlmeyer JL, Meyerholz DK, Tanas MR, et al. A porcine model of neurofibromatosis type 1 that mimics the human disease. *JCI Insight.* 2018;3(12):e120402.
111. Mautner VF, Kluwe L, Thakker SD, Lark RA. Treatment of ADHD in neurofibromatosis type 1. *Dev Med Child Neurol.* 2002;44(3):164–70.
112. Soucy EA, Gao F, Gutmann DH, Dunn CM. Developmental delays in children with neurofibromatosis type 1. *J Child Neurol.* 2012;27(5):641–4.
113. Ferner RE, Hughes RAC, Weinman J. Intellectual impairment in neurofibromatosis 1. *J Neurol Sci.* 1996;138(1):125–33.
114. Morris SM, Acosta MT, Garg S, Green J, Huson S, Legius E, et al. Disease Burden and Symptom Structure of Autism in Neurofibromatosis Type 1: a study of the international NF1-ASD Consortium Team (INFACT). *JAMA Psychiatry.* 2016;73:1276–84.
115. Plasschaert E, Van Eylen L, Descheemaeker M-J, Noens I, Legius E, Steyaert J. Executive functioning deficits in children with neurofibromatosis type 1: The influence of intellectual and social functioning. *Am J Med Genet B Neuropsychiatr Genet.* 2016;171(3):348–62.
116. Plasschaert E, Descheemaeker MJ, Van Eylen L, Noens I, Steyaert J, Legius E. Prevalence of Autism Spectrum Disorder symptoms in children with neurofibromatosis type 1. *Am J Med Genet B Neuropsychiatr Genet.* 2015;168b(1):72–80.
117. Garg S, Green J, Leadbitter K, Emsley R, Lehtonen A, Evans DG, et al. Neurofibromatosis type 1 and autism spectrum disorder. *Pediatrics.* 2013;132(6):e1642–8.
118. Walsh KS, Velez JI, Kardel PG, Imas DM, Muenke M, Packer RJ, et al. Symptomatology of autism spectrum disorder in a population with neurofibromatosis type 1. *Dev Med Child Neurol.* 2013;55(2):131–8.
119. Chatterjee N, Perrimon N. What fuels the fly: energy metabolism in *Drosophila* and its application to the study of obesity and diabetes. *Sci Adv.* 2021;7(24):eabg4336.
120. Musselman LP, Kühnlein RP. *Drosophila* as a model to study obesity and metabolic disease. *J Exp Biol.* 2018;221(Suppl_1):jeb163881.
121. Gatto CL, Brodie K. *Drosophila* modeling of heritable neurodevelopmental disorders. *Curr Opin Neurobiol.* 2011;21(6):834–41.
122. Williams JA, Su HS, Bernards A, Field J, Sehgal A. A circadian output in *Drosophila* mediated by neurofibromatosis-1 and Ras/MAPK. *Science.* 2001;293(5538):2251–6.
123. Machado Almeida P, Lago Solis B, Stickley L, Feidler A, Nagoshi E. Neurofibromin 1 in mushroom body neurons mediates circadian wake drive through activating cAMP–PKA signaling. *Nat Commun.* 2021;12(1):5758.
124. Bai L, Sehgal A. Anaplastic Lymphoma Kinase Acts in the *Drosophila* Mushroom Body to Negatively Regulate Sleep. *PLoS Genet.* 2015;11(11):e1005611.
125. Buchanan ME, Davis RL. A distinct set of *Drosophila* brain neurons required for neurofibromatosis type 1-dependent learning and memory. *J Neurosci.* 2010;30(30):10135–43.
126. Ho IS, Hannan F, Guo HF, Hakker I, Zhong Y. Distinct functional domains of neurofibromatosis type 1 regulate immediate versus long-term memory formation. *J Neurosci.* 2007;27(25):6852–7.
127. King LB, Boto T, Botero V, Aviles AM, Jomsky BM, Joseph C, et al. Developmental loss of neurofibromin across distributed neuronal circuits drives excessive grooming in *Drosophila*. *PLoS Genet.* 2020;16(7):e1008920.
128. King LB, Koch M, Murphy KR, Velazquez Y, Ja WW, Tomchik SM. Neurofibromin loss of function drives excessive grooming in *drosophila*. *G3 (Bethesda).* 2016;6(4):1083–93.
129. Brown EB, Zhang J, Lloyd E, Lanzon E, Botero V, Tomchik S, et al. Neurofibromin 1 mediates sleep depth in *Drosophila*. *PLoS Genet.* 2023;19(12):e1011049.
130. Moscato EH, Dubowy C, Walker JA, Kayser MS. Social behavioral deficits with loss of Neurofibromin emerge from peripheral chemosensory neuron dysfunction. *Cell Rep.* 2020;32(1):107856.
131. Dyson A, Ryan M, Garg S, Evans DG, Baines RA. Loss of NF1 in *drosophila* larvae causes tactile hypersensitivity and impaired synaptic transmission at the neuromuscular junction. *J Neurosci.* 2022;42(50):9450–72.
132. Dyson A, Ryan M, Garg S, Evans DG, Baines RA. A Targeted, Low-Throughput Compound Screen in a *Drosophila* Model of Neurofibromatosis Type 1 Identifies Simvastatin and BMS-204352 as Potential Therapies for Autism Spectrum Disorder (ASD). *Eneuro.* 2023;10(5):ENEURO.0461–ENEURO.0461.
133. Oliveira AF, Yasuda R. Neurofibromin is the major ras inactivator in dendritic spines. *J Neurosci.* 2014;34(3):776–83.
134. Li W, Cui Y, Kushner SA, Brown RA, Jentsch JD, Frankland PW, et al. The HMG-CoA reductase inhibitor lovastatin reverses the learning and attention deficits in a mouse model of neurofibromatosis type 1. *Curr Biol.* 2005;15(21):1961–7.
135. Guilding C, McNair K, Stone TW, Morris BJ. Restored plasticity in a mouse model of neurofibromatosis type 1 via inhibition of hyperactive ERK and CREB. *Eur J Neurosci.* 2007;25(1):99–105.
136. Silva AJ, Frankland PW, Marowitz Z, Friedman E, Laszlo GS, Cioffi D, et al. A mouse model for the learning and memory deficits associated with neurofibromatosis type I. *Nat Genet.* 1997;15(3):281–4.
137. Hingtgen CM. Neurofibromatosis: the role of guanosine triphosphatase activating proteins in sensory neuron function. *Sheng Li Xue Bao.* 2008;60(5):581–3.
138. Diggs-Andrews KA, Tokuda K, Izumi Y, Zorunski CF, Wozniak DF, Gutmann DH. Dopamine deficiency underlies learning deficits in neurofibromatosis-1 mice. *Ann Neurol.* 2013;73(2):309–15.
139. Ryu HH, Kang M, Park J, Park SH, Lee YS. Enriched expression of NF1 in inhibitory neurons in both mouse and human brain. *Mol Brain.* 2019;12(1):60.

140. Friedman JM, Riccardi VM. Neurofibromatosis : phenotype, natural history, and pathogenesis. 3rd ed. Baltimore: Johns Hopkins University Press Baltimore; 1999.
141. Souza M, Jansen A, Martins A, Rodrigues L, Rezende N. Body composition in adults with neurofibromatosis type 1. *Rev Assoc Med Bras* (1992). 2016;62(9):831–6.
142. Koga M, Yoshida Y, Imafuku S. Nutritional, muscular and metabolic characteristics in patients with neurofibromatosis type 1. *J Dermatol*. 2016;43(7):799–803.
143. Ferrara UP, Tortora C, Rosano C, Assunto A, Rossi A, Pagano S, et al. Bone metabolism in patients with type 1 neurofibromatosis: key role of sun exposure and physical activity. *Sci Rep*. 2022;12(1):4368.
144. Soucy EA, van Oppen D, Nejedly NL, Gao F, Gutmann DH, Hollander AS. Height assessments in children with neurofibromatosis type 1. *J Child Neurol*. 2013;28(3):303–7.
145. Souza JF, Passos RL, Guedes AC, Rezende NA, Rodrigues LO. Muscular force is reduced in neurofibromatosis type 1. *J Musculoskelet Neuronal Interact*. 2009;9(1):15–7.
146. Johnson BA, MacWilliams BA, Carey JC, Viskochil DH, D'Astous JL, Stevenson DA. Motor proficiency in children with neurofibromatosis type 1. *Pediatr Phys Ther*. 2010;22(4):344–8.
147. Martins AS, Jansen AK, Rodrigues LO, Matos CM, Souza ML, de Souza JF, et al. Lower fasting blood glucose in neurofibromatosis type 1. *Endocr Connect*. 2016;5(1):28–33.
148. Martins AS, Jansen AK, Rodrigues LOC, Matos CM, Souza MLR, Miranda DM, et al. Increased insulin sensitivity in individuals with neurofibromatosis type 1. *Arch Endocrinol Metab*. 2018;62(1):41–6.
149. Ozhan B, Ozguven AA, Ersoy B. Neurofibromatosis type 1 and diabetes mellitus: an unusual association. *Case Rep Endocrinol*. 2013;2013:689107.
150. Kallionpää RA, Peltonen S, Leppävirta J, Pöyhönen M, Auranen K, Järveläinen H, et al. Haploinsufficiency of the NF1 gene is associated with protection against diabetes. *J Med Genet*. 2021;58(6):378–84.
151. Vassallo G, Mughal Z, Robinson L, Weisberg D, Roberts SA, Hupton E, et al. Perceived fatigue in children and young adults with neurofibromatosis type 1. *J Paediatr Child Health*. 2020;56(6):878–83.
152. Gehart H, Kumpf S, Iltner A, Ricci R. MAPK signalling in cellular metabolism: stress or wellness? *EMBO Rep*. 2010;11(11):834–40.
153. Guo YJ, Pan WW, Liu SB, Shen ZF, Xu Y, Hu LL. ERK/MAPK signalling pathway and tumorigenesis (Review). *Exp Ther Med*. 2020;19(3):1997–2007.
154. Masgras I, Ciscato F, Brunati AM, Tibaldi E, Indraccolo S, Curtarello M, et al. Absence of Neurofibromin Induces an Oncogenic Metabolic Switch via Mitochondrial ERK-Mediated Phosphorylation of the Chaperone TRAP1. *Cell Rep*. 2017;18(3):659–72.
155. Tritz R, Benson T, Harris V, Hudson FZ, Mintz J, Zhang H, et al. Nf1 heterozygous mice recapitulate the anthropometric and metabolic features of human neurofibromatosis type 1. *Transl Res*. 2021;228:52–63.
156. Sullivan K, El-Hoss J, Quinlan KG, Deo N, Garton F, Seto JT, et al. NF1 is a critical regulator of muscle development and metabolism. *Hum Mol Genet*. 2014;23(5):1250–9.
157. Brannan CI, Perkins AS, Vogel KS, Ratner N, Nordlund ML, Reid SW, et al. Targeted disruption of the neurofibromatosis type-1 gene leads to developmental abnormalities in heart and various neural crest-derived tissues. *Genes Dev*. 1994;8(9):1019–29.
158. Summers MA, Rupasinghe T, Vasiljevski ER, Evesson FJ, Mikulec K, Peacock L, et al. Dietary intervention rescues myopathy associated with neurofibromatosis type 1. *Hum Mol Genet*. 2018;27(4):577–88.
159. Summers MA, Vasiljevski ER, Mikulec K, Peacock L, Little DG, Schindeler A. Developmental dosing with a MEK inhibitor (PD0325901) rescues myopathic features of the muscle-specific but not limb-specific Nf1 knockout mouse. *Mol Genet Metab*. 2018;123(4):518–25.
160. Wei X, Franke J, Ost M, Wardelmann K, Borno S, Timmermann B, et al. Cell autonomous requirement of neurofibromin (Nf1) for postnatal muscle hypertrophic growth and metabolic homeostasis. *J Cachexia Sarcopenia Muscle*. 2020;11(6):1758–78.
161. Walker JA, Tchoudakova AV, McKenney PT, Brill S, Wu D, Cowley GS, et al. Reduced growth of *Drosophila* neurofibromatosis 1 mutants reflects a non-cell-autonomous requirement for GTPase-Activating Protein activity in larval neurons. *Genes Dev*. 2006;20(23):3311–23.
162. Hannan F, Ho I, Tong JJ, Zhu Y, Nurnberg P, Zhong Y. Effect of neurofibromatosis type I mutations on a novel pathway for adenylyl cyclase activation requiring neurofibromin and Ras. *Hum Mol Genet*. 2006;15(7):1087–98.
163. Tong JJ, Schriener SE, McCleary D, Day BJ, Wallace DC. Life extension through neurofibromin mitochondrial regulation and antioxidant therapy for neurofibromatosis-1 in *Drosophila melanogaster*. *Nat Genet*. 2007;39(4):476–85.
164. Botero V, Stanhope BA, Brown EB, Greci EC, Boto T, Park SJ, et al. Neurofibromin regulates metabolic rate via neuronal mechanisms in *Drosophila*. *Nat Commun*. 2021;12(1):4285.
165. Maurer GW, Malita A, Nagy S, Koyama T, Werge TM, Halberg KA, et al. Analysis of genes within the schizophrenia-linked 22q11.2 deletion identifies interaction of night owl/LZTR1 and NF1 in GABAergic sleep control. *PLoS genetics*. 2020;16(4):e1008727.
166. DeBella K, Szudek J, Friedman JM. Use of the National Institutes of Health Criteria for Diagnosis of Neurofibromatosis 1 in Children. *Pediatriatrics*. 2000;105(3):608–14.
167. De Schepper S, Maertens O, Callens T, Naeyaert J-M, Lambert J, Messiaen L. Somatic Mutation Analysis in NF1 Cafe au lait Spots Reveals Two NF1 Hits in the Melanocytes. *J Invest Dermatol*. 2008;128(4):1050–3.
168. Lubs MLE, Bauer MS, Formas ME, Djokic B. Lisch Nodules in Neurofibromatosis Type 1. *N Engl J Med*. 1991;324(18):1264–6.
169. Stevenson DA, Birch PH, Friedman JM, Viskochil DH, Balestrazzi P, Boni S, et al. Descriptive analysis of tibial pseudarthrosis in patients with neurofibromatosis 1. *Am J Med Genet*. 1999;84(5):413–9.
170. Kjell VR, Hilde B, Eric L, Johan L, Armand L. Prevalence of neurofibromatosis type 1 in congenital pseudarthrosis of the tibia. *Eur J Pediatr*. 2016;175(9):1193–8.
171. Jackson IT, Carbonnel A, Potparic Z, Shaw K. Orbitotemporal neurofibromatosis: classification and treatment. *Plast Reconstr Surg*. 1993;92(1):1–11.
172. Brunetti-Pierrri N, Doty SB, Hicks J, Phan K, Mendoza-Londono R, Blazo M, et al. Generalized metabolic bone disease in Neurofibromatosis type I. *Mol Genet Metab*. 2008;94(1):105–11.
173. Crawford AH, Herrera-Soto J. Scoliosis Associated with Neurofibromatosis. *Orthop Clin North Am*. 2007;38(4):553–62.
174. Szudek J, Birch P, Friedman JM. Growth in North American white children with neurofibromatosis 1 (NF1). *J Med Genet*. 2000;37(12):933–8.
175. Zessis NR, Gao F, Vadlamudi G, Gutmann DH, Hollander AS. Height growth impairment in children with neurofibromatosis type 1 is characterized by decreased pubertal growth velocity in both sexes. *J Child Neurol*. 2018;33(12):762–6.
176. Stevenson DA, Yan J, He Y, Li H, Liu Y, Zhang Q, et al. Multiple increased osteoclast functions in individuals with neurofibromatosis type 1. *Am J Med Genet A*. 2011;155a(5):1050–9.
177. Caffarelli C, Gonnelli S, Tanzilli L, Vivarelli R, Tamburello S, Balestri P, et al. Quantitative ultrasound and dual-energy x-ray absorptiometry in children and adolescents with neurofibromatosis of type 1. *J Clin Densitom*. 2010;13(1):77–83.
178. Lammert M, Kappler M, Mautner VF, Lammert K, Störkel S, Friedman JM, et al. Decreased bone mineral density in patients with neurofibromatosis 1. *Osteoporos Int*. 2005;16(9):1161–6.
179. Kolanczyk M, Kossler N, Kühnisch J, Lavitas L, Stricker S, Wilkening U, et al. Multiple roles for neurofibromin in skeletal development and growth. *Hum Mol Genet*. 2007;16(8):874–86.
180. Paria N, Khalid A, Shen B, Lemoine B, Chan J, Kidane YH, et al. Molecular dissection of somatic skeletal disease in neurofibromatosis type 1. *J Bone Miner Res*. 2022;38(2):288–99.
181. Wang W, Nyman JS, Ono K, Stevenson DA, Yang X, Eleftheriou F. Mice lacking Nf1 in osteochondroprogenitor cells display skeletal dysplasia similar to patients with neurofibromatosis type I. *Hum Mol Genet*. 2011;20(20):3910–24.
182. Uthoff J, Larson J, Sato TS, Hammond E, Schroeder KE, Rohret F, et al. Longitudinal phenotype development in a minipig model of neurofibromatosis type 1. *Sci Rep*. 2020;10(1):5046.
183. Wang S, Mautner VF, Buchert R, Flibotte S, Suppa P, Friedman JM, et al. Alterations in brain morphology by MRI in adults with neurofibromatosis 1. *Orphanet J Rare Dis*. 2021;16(1):462.
184. Cutting LE, Cooper KL, Koth CW, Mostofsky SH, Kates WR, Denckla MB, et al. Megalencephaly in NF1: predominantly white matter contribution and mitigation by ADHD. *Neurology*. 2002;59(9):1388–94.

185. Greenwood RS, Tupler LA, Whitt JK, Buu A, Dombeck CB, Harp AG, et al. Brain morphometry, T2-weighted hyperintensities, and IQ in children with neurofibromatosis type 1. *Arch Neurol*. 2005;62(12):1904–8.
186. Moore BD 3rd, Slopis JM, Jackson EF, De Winter AE, Leeds NE. Brain volume in children with neurofibromatosis type 1: relation to neuropsychological status. *Neurology*. 2000;54(4):914–20.
187. Steen RG, Taylor JS, Langston JW, Glass JO, Brewer VR, Reddick WE, et al. Prospective evaluation of the brain in asymptomatic children with neurofibromatosis type 1: relationship of macrocephaly to T1 relaxation changes and structural brain abnormalities. *AJNR Am J Neuroradiol*. 2001;22(5):810–7.
188. Karlsgodt KH, Rosser T, Lutkenhoff ES, Cannon TD, Silva A, Bearden CE. Alterations in white matter microstructure in neurofibromatosis-1. *PLoS One*. 2012;7(10):e47854.
189. Said SM, Yeh TL, Greenwood RS, Whitt JK, Tupler LA, Krishnan KR. MRI morphometric analysis and neuropsychological function in patients with neurofibromatosis. *NeuroReport*. 1996;7(12):1941–4.
190. Dubovsky EC, Booth TN, Vezina G, Samango-Sprouse CA, Palmer KM, Brasseux CO. MR imaging of the corpus callosum in pediatric patients with neurofibromatosis type 1. *AJNR Am J Neuroradiol*. 2001;22(1):190–5.
191. Wignall EL, Griffiths PD, Papadakis NG, Wilkinson ID, Wallis LI, Bandmann O, et al. Corpus callosum morphology and microstructure assessed using structural MR imaging and diffusion tensor imaging: initial findings in adults with neurofibromatosis type 1. *AJNR Am J Neuroradiol*. 2010;31(5):856–61.
192. DiMario FJ Jr, Ramsby G, Greenstein R, Langshur S, Dunham B. Neurofibromatosis type 1: magnetic resonance imaging findings. *J Child Neurol*. 1993;8(1):32–9.
193. Ji J, Shimony J, Gao F, McKinstry RC, Gutmann DH. Optic nerve tortuosity in children with neurofibromatosis type 1. *Pediatr Radiol*. 2013;43(10):1336–43.
194. Wang S, Friedman JM, Suppa P, Buchert R, Mautner VF. White matter is increased in the brains of adults with neurofibromatosis 1. *Orphanet J Rare Dis*. 2022;17(1):115.
195. Kayl AE, Moore BD 3rd, Slopis JM, Jackson EF, Leeds NE. Quantitative morphology of the corpus callosum in children with neurofibromatosis and attention-deficit hyperactivity disorder. *J Child Neurol*. 2000;15(2):90–6.
196. Aydin S, Kurtcan S, Alkan A, Guler S, Filiz M, Yilmaz TF, et al. Relationship between the corpus callosum and neurocognitive disabilities in children with NF-1: diffusion tensor imaging features. *Clin Imaging*. 2016;40(6):1092–5.
197. Shofty B, Bergmann E, Zur G, Asleh J, Bosak N, Kavushansky A, et al. Autism-associated Nf1 deficiency disrupts corticocortical and corticostriatal functional connectivity in human and mouse. *Neurobiol Dis*. 2019;130:104479.
198. Tomson SN, Schreiner MJ, Narayan M, Rosser T, Enrique N, Silva AJ, et al. Resting state functional MRI reveals abnormal network connectivity in neurofibromatosis 1. *Hum Brain Mapp*. 2015;36(11):4566–81.
199. Loitfelder M, Huijbregts SC, Veer IM, Swaab HS, Van Buchem MA, Schmidt R, et al. Functional Connectivity Changes and Executive and Social Problems in Neurofibromatosis Type I. *Brain Connect*. 2015;5(5):312–20.
200. Baudou E, Nemmi F, Peran P, Cignetti F, Blais M, Maziero S, et al. Atypical connectivity in the cortico-striatal network in NF1 children and its relationship with procedural perceptual-motor learning and motor skills. *J Neurodev Disord*. 2022;14(1):15.
201. Ibrahim AFA, Montojo CA, Haut KM, Karlsgodt KH, Hansen L, Congdon E, et al. Spatial working memory in neurofibromatosis 1: Altered neural activity and functional connectivity. *Neuroimage Clin*. 2017;15:801–11.
202. Baudou E, Nemmi F, Biotteau M, Maziero S, Peran P, Chaix Y. Can the Cognitive Phenotype in Neurofibromatosis Type 1 (NF1) Be Explained by Neuroimaging? A Review *Front Neurol*. 2019;10:1373.
203. Mennigen E, Schuette P, Vajdi A, Pacheco L, Rosser T, Bearden CE. Reduced higher dimensional temporal dynamism in neurofibromatosis type 1. *Neuroimage Clin*. 2019;22:101692.
204. Chabernaud C, Mennes M, Kardel PG, Gaillard WD, Kalbfleisch ML, Vanmeter JW, et al. Lovastatin regulates brain spontaneous low-frequency brain activity in neurofibromatosis type 1. *Neurosci Lett*. 2012;515(1):28–33.
205. Nemmi F, Cignetti F, Assaiante C, Maziero S, Audic F, Peran P, et al. Discriminating between neurofibromatosis-1 and typically developing children by means of multimodal MRI and multivariate analyses. *Hum Brain Mapp*. 2019;40(12):3508–21.
206. Wang Y, Kim E, Wang X, Novitch Bennett G, Yoshikawa K, Chang L-S, et al. ERK Inhibition Rescues Defects in Fate Specification of Nf1-Deficient Neural Progenitors and Brain Abnormalities. *Cell*. 2012;150(4):816–30.
207. Lakkis MM, Golden JA, O'Shea KS, Epstein JA. Neurofibromin deficiency in mice causes Exencephaly and is a modifier for splotch neural tube defects. *Dev Biol*. 1999;212(1):80–92.
208. Gutmann DH, Loehr A, Zhang Y, Kim J, Henkemeyer M, Cashen A. Haploinsufficiency for the neurofibromatosis 1 (NF1) tumor suppressor results in increased astrocyte proliferation. *Oncogene*. 1999;18(31):4450–9.
209. Gutmann DH, Wu YL, Hedrick NM, Zhu Y, Guha A, Parada LF. Heterozygosity for the neurofibromatosis 1 (NF1) tumor suppressor results in abnormalities in cell attachment, spreading and motility in astrocytes. *Hum Mol Genet*. 2001;10(26):3009–16.
210. Zhu Y, Romero MI, Ghosh P, Ye Z, Charnay P, Rushing EJ, et al. Ablation of NF1 function in neurons induces abnormal development of cerebral cortex and reactive gliosis in the brain. *Genes Dev*. 2001;15(7):859–76.
211. Zhu Y, Harada T, Liu L, Lush ME, Guignard F, Harada C, et al. Inactivation of NF1 in CNS causes increased glial progenitor proliferation and optic glioma formation. *Development*. 2005;132(24):5577–88.
212. Hegedus B, Dasgupta B, Shin JE, Emmett RJ, Hart-Mahon EK, Elghazi L, et al. Neurofibromatosis-1 Regulates Neuronal and Glial Cell Differentiation from Neuroglial Progenitors In Vivo by Both cAMP- and Ras-Dependent Mechanisms. *Cell Stem Cell*. 2007;1(4):443–57.
213. Hegedus B, Yeh TH, Lee DY, Emmett RJ, Li J, Gutmann DH. Neurofibromin regulates somatic growth through the hypothalamic-pituitary axis. *Hum Mol Genet*. 2008;17(19):2956–66.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.